# HiFi II M-MLV(H-) Reverse Transcriptase

Project No. H665664 (10,000 U) Storage condition: -20°C storage.

#### Product content

individual parts making up a compound	H665664 10000 U
HiFi II M-MLV(H-) (200U /μL)	50 μL
5×SuperRT Buffer	1 mL

#### Product Introduction

HiFi II M-MLV(H-) is a mutated M-MLV gene recombinant and expressed in E. coli engineered bacteria as a reverse transcriptase that catalyzes complementary DNA polymerization reactions using RNA or DNA:RNA hybrid strands as templates. The mutated HiFi II M-MLV(H-) reverse transcriptase is deficient in RNase H activity, which reduces the degradation of RNA in the reverse transcription reaction and makes it easier to obtain full-length cDNAs. HiFi II M-MLV(H-) reverse transcriptase is able to synthesize the first strand of cDNAs at 55° C, providing higher specificity, stability, and the ability to synthesize cDNAs of up to 12 kb with high cDNA yields. High cDNA yield. It is suitable for first-strand cDNA synthesis, RT-PCR, RT-qPCR and full-length cDNA library construction.

## Active Definition

The amount of enzyme required to catalyze the incorporation of 1 nmol of dTTP in 10 min at 37° C is defined as one unit of activity (U), using Poly (A) as a template and oligo (dT) as a primer.

# Quality control

200 U of this enzyme and 1  $\mu$ g of 16 S, 23 S rRNA were reacted at 37° C for 1 h. The electrophoretic bands of RNA did not change.

#### Caveat

- 1. RNase contamination should be avoided during operation to prevent RNA degradation or cross-contamination in the experiment. It is recommended that RNA operation be carried out in a special area, with special instruments and consumables, and that operators wear masks and disposable gloves and change gloves frequently.
- 2. Use disposable plastic containers as much as possible for the experiment. If glassware is used, it should be treated with 0.1% DEPC (diethyl ether pyrocarbonate) aqueous solution at 37°C for 12 hours and autoclaved at 120°C for 30 minutes before use, or the glassware should be sterilized by dry heat at 180°C for 60 minutes before use. Sterile water used in experiments should be treated with 0.1% DEPC and autoclaved.
- 3. All reagents in this kit should be mixed gently, upside down, to avoid foaming and centrifuged briefly before use. The enzymes should be returned to  $-20^{\circ}$ C as soon as possible after use to avoid repeated freezing and thawing.
- 4. If the amount of starting RNA is less than 50 ng, it is recommended to add RNAase inhibitor (RNasin). This kit is not supplied.

## Usage

Note: 10 ng-5  $\mu$ g of total RNA creates a 20  $\mu$ L reaction system, if the amount of total RNA is greater than 5  $\mu$ g, please scale up the reaction system proportionally.

#### i Reverse transcription procedure:

- 1. Dissolve RNA template, primers, dNTP Mix, SuperRT Buffer, HiFi II M-MLV(H-) and RNase-Free Water and set aside on ice.
- 2. Prepare the reaction system according to the following table in a total volume of 20  $\,\mu\,L.$

reagents	20 μl reaction system	final concentration
dNTP Mix, 2.5 mM Each	4 μ1	500 μM Each
Oligo-dT Primer, 100 μ M or Random Primers , 50 μ M or Specific Primer , 10 μ M	1 μ1	/
RNA Template	Χ μ1	1 ng-5 μg
5×SuperRT Buffer	4 μ1	1 ×

HiFi II M-MLV(H-) (200U /μL)	0.5-1 μL	/
RNase-Free Water	up to 20 µL	/

**Note:** If the amount of starting RNA is less than 50ng, it is recommended to add RNAase inhibitor (RNasin). This kit is not supplied.

- 3. Mix by vortex shaking and centrifuge briefly so that the solution on the walls of the tube collects at the bottom.
- 4. Incubate at  $55^{\circ}$  C for 1-30 minutes and at  $85^{\circ}$  C for 5 minutes. At the end of the reaction, centrifuge briefly and place on ice to cool.
- 5. The reverse transcription product can be used directly in PCR and fluorescence quantitative PCR reactions, or placed at  $-20^{\circ}$ C for long-term storage.

# ii The following steps are recommended if reverse transcription efficiency is low, or if the RNA template secondary structure is complex and GC content is high:

- 1. Dissolve RNA template, primers, dNTP Mix, SuperRT Buffer, HiFi II M-MLV(H-) and RNase-Free Water and set aside on ice.
- 2. Prepare the reaction system according to the following table in a total volume of 15  $\,\mu\,L$  .

reagents	20 μl reaction system	final concentration
dNTP Mix, 2.5 mM Each	4 μ1	500 μM Each
Oligo-dT Primer, 100 μ M or Random Primers , 50 μ M or Specific Primer , 10 μ M	1 μ1	/
RNA Template	Χ μ1	1 ng-5 μg
RNase-Free Water	up to 15 µL	/

- 3. Incubate at  $70^{\circ}$  C for 10 minutes and rapidly ice bath for 2 minutes.
- 4. Centrifuge briefly so that the solution on the walls of the tube collects at the bottom.
- 5. Add 4 µL of 5×SuperRT Buffer to the above reaction solution.

**Note:** If the amount of starting RNA is less than 50 ng, it is recommended that an RNA inhibitor (RNasin) be added. It is not supplied with this kit.

- 6. Blow gently to mix, if the reverse transcription primer is an Oligo-dT Primer or Specific Primer.
- 7. Incubate at 42° C for 2 minutes or 25° C for 10 minutes if the reverse transcription primers are Random Primers.

- 8. Add 1  $\mu$ L of HiFi II M-MLV(H-) (200 U/ $\mu$ L) and mix by gentle pipetting. Incubate at 55° C for 50 minutes. Incubate at 85° C for 5 minutes. At the end of the reaction, centrifuge briefly and place on ice to cool.
- 9. Reverse transcription products can be used directly in PCR reactions and fluorescence quantitative PCR reactions, or placed at  $-20^{\circ}$  C for long-term storage.